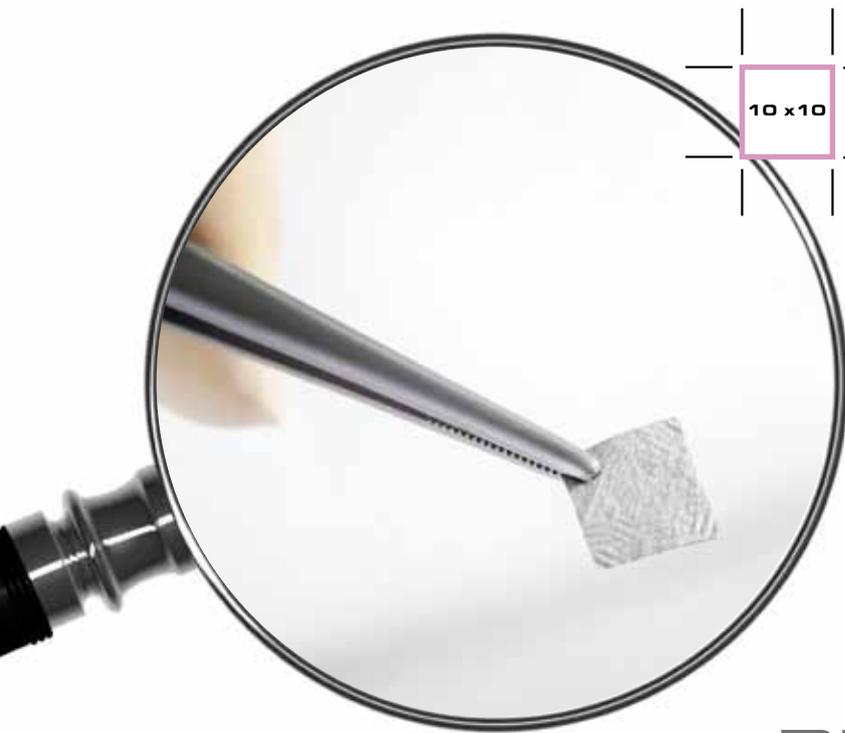


SMARTBRANE

Resorbable Pericardium Membrane



10 x 10

NEW!
THE SMALLEST
MEMBRANE
10 x 10 mm

SIMPLE

RELIABLE

PURE

MORE
ECONOMIC



**NEW
MINI!**



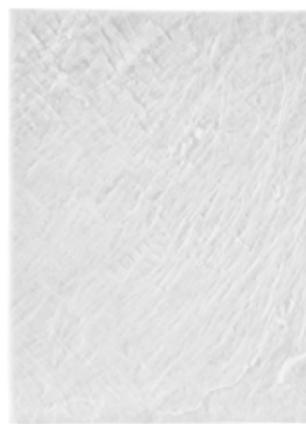
10 x 10 mm



15 x 20 mm



20 x 30 mm



30 x 40 mm

MORE ECONOMIC

MORE ECONOMIC

The smallest membrane 10 x 10 mm

SMARTBRANE is a resorbable collagen membrane made of porcine pericardium. Thus, it features all benefits of a modern native collagen membrane.

In addition to the standard membrane sizes, it is available in a mini format of 10 x 10 mm. This offers a more economic membrane solution especially for regeneration of small bone defects to optimize your cost-benefit structure.



NEW!
**THE SMALLEST
MEMBRANE**
10 x 10 mm

SIMPLE

SIMPLE

Optimized handling properties ensuring straight-forward application

The supercritical carbon dioxide (scCO₂) cleaning process is gently removing unwanted materials (e.g. cells, lipids) while preserving the natural collagen matrix as well as the natural crosslinking of the collagen fibers.^{1,2}

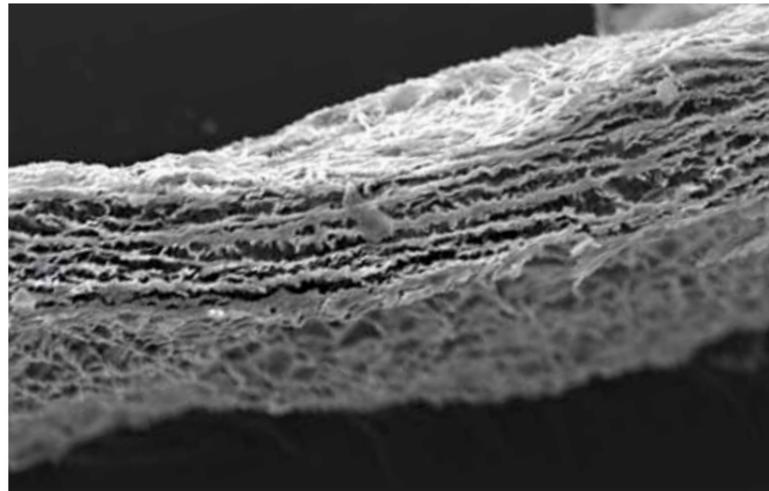
As a result, SMARTBRANE is characterized by optimal material stability as the biomechanical characteristics of porcine pericardium tissue are preserved.³

SMARTBRANE...

- features an adequate tensile strength
- is very adaptable to bony surfaces without sticking to the graft or instrument
- is very thin (<0,4mm) for facilitated augmentation and wound closure



SMARTBRANE rehydrated: Excellent adaptation to surfaces without sticking to graft or instrument.



SMARTBRANE cross-section (magnification x 40) featuring intact structure as well as a natural interconnective porous system.

PURE

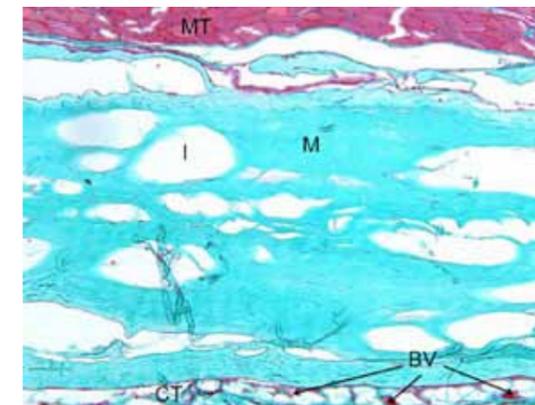
Excellent biocompatibility for improved wound healing

SMARTBRANE is manufactured using an innovative and highly effective cleaning technology based on supercritical carbon dioxide (scCO₂).

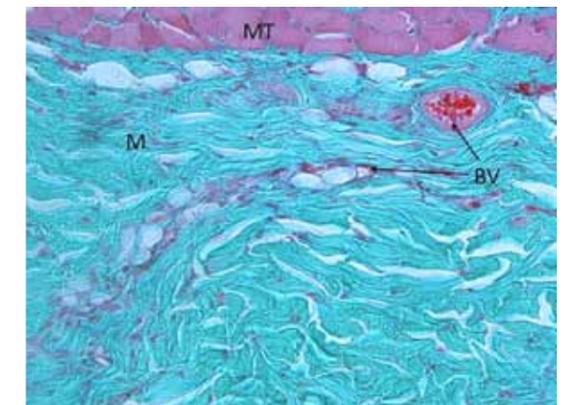
This process results in a higher purity and creates a biocompatible base for immediate new bone ingrowth.^{1,2}

It provides highest possible biocompatibility characteristics due to its porcine origin and the scCO₂ cleaning process.

Histological examination in vivo⁴



After 1 week of subcutaneous implantation in a rat muscle: SMARTBRANE (M) is already connected to the muscular tissue (MT), no signs of inflammatory reactions.



After 2 weeks the first blood vessels (BV) are invading SMARTBRANE (M), no signs of inflammatory reactions.

RELIABLE

RELIABLE

Natural collagen matrix preserved by scCO₂ cleaning technology for enhanced graft performance

SMARTBRANE is made of porcine pericardium and thus presents optimal matrix composition and a natural dense 3D-network collagen structure optimally preserved after scCO₂ purification.

The preserved natural collagen matrix plays an important role for blood clotting and promotes cell attachment.⁵

The membrane has a resorption time of 8-12 weeks providing adequate barrier function for usage in standard GBR cases.⁶



CASE REPORT

CASE REPORT

Augmentation of a dehiscence-type defect around dental implant



Surgery
Dehiscence defect around bone level implant.



Augmentation with Xenograft bone.



Coverage of bone graft material with SMARTBRANE – the membrane can be easily positioned and is adapting ideally to the defect geometry.

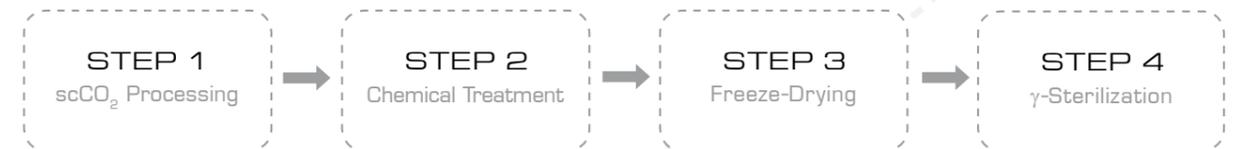


Suture removal
Optimal initial healing pattern: no signs of irritation.

TECHNOLOGY

TECHNOLOGY

scCO₂ cleaning process as basis for optimal matrix properties and maximal graft safety



STEP 1

Supercritical Carbon Dioxide (scCO₂) Processing

- Carbon dioxide is in its supercritical state when both the temperature and pressure equal or exceed the critical point of 31°C and 73 atm.
- In this supercritical state, CO₂ has both gas-like and liquid-like qualities.
- By its effective tissue perfusion and removing capabilities of unwanted substances it provides ideal conditions for cleaning and sterilising tissues.^{1,2}
- Furthermore, scCO₂ is known to be highly efficient against all kinds of pathogens.⁷

Step 2

Chemical Treatment

- Various chemical treatment steps are applied to provide a pure membrane matrix by inactivating and removing residual non-collagenous proteins and enzymes. This results in a further increased safety level for pathogen inactivation.⁸

Step 3

Freeze-Drying

- Freeze-drying allows gentle preservation, retaining the original 3D structure of the xenograft.
- After freeze-drying, products can be stored at room temperature and generally have a longer shelf-life.

Step 4

Gamma-Sterilization

- The combination of scCO₂ cleaning process and terminal gamma-sterilization provides highest possible viral and bacterial inactivation and results in a sterile (SAL > 10⁻⁶) and highly biocompatible bone graft.^{1,9}



REFERENCES

1. Nichols A, Burns DC, Christopher R. Studies on the Sterilization of Human Bone and Tendon Musculoskeletal Allograft Tissue Using Supercritical Carbon Dioxide. *Journal of Orthopaedics* 2009.
2. Sawada K, Terada D, Yamaoka T, Kitamura S, Fujisato T. Cell removal with supercritical carbon dioxide for acellular artificial tissue. *J Chemical Technol Biotechnol* 2008;83(6):943–949.
3. Internal testing results, data on file.
4. SMARTBRANE subcutaneous implantation test, data on file.
5. Brett D. A Review of Collagen and Collagen-based Wound Dressings. *Wounds* 2008;20(12).
6. Internal testing results, data on file.
7. a. Pages F, Poirier B, Barbier Y, Frayssinet P, Joffret M-L, Majewski W, Bonel G, Larzul D. Viral Inactivation of Human Bone Tissue using supercritical Fluid Extraction. *ASAIO Journal* 1998;44:289-293. 7b. Qiu QQ, Leamy P, Brittingham J, Pomerleau J, Kabaria N, Connor J. Inactivation of bacterial spores and viruses in biological material using supercritical carbon dioxide with sterilant. *J Biomed Mater Res B Appl Biomater*: 2009;91(2):572-8. 7c. Dillow AK, Dehghani F, Hrkach JS, Foster NR, Langer R. Bacterial inactivation by using near- and supercritical carbon dioxide. *Proc Natl Acad Sci U S A*. 1999;96(18):10344-8.
8. Sofer G, Lister DC, Boose JA. Part 6, Inactivation Methods Grouped by Virus. *BioPharmInternational* 2003;6 Supplement:S37-S42.
9. Thomas FC, Ouwerkerk T, McKercher P. Inactivation by gamma irradiation of animal viruses in simulated laboratory effluent. *Appl Environ Microbiol*. 1982;43(5):1051–1056.

Clinical pictures by courtesy Dr. Kai Fischer (Germany).

Manufactured by REGEDENT AG, Zollikerstrasse 144, CH - 8008 Zürich

CE0086

Version 150818

■ CONTACT

REGEDENT AG
Zollikerstrasse 144
CH - 8008 Zürich
Tel +41 (0) 44 - 7 00 37 77
Fax +41 (0) 44 - 7 00 47 97

REGEDENT GmbH
Pfarrgasse 6
D - 97337 Dettelbach
Tel +49 (0) 93 24 - 6 04 99 27
Fax +49 (0) 93 24 - 6 04 99 26

Mail info@regedent.com
www.regedent.com

